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Research Article PHOSPHORUS BIOFERTILIZER AS EFFICIENT FERTILIZER FOR IMPROVING NUTRIENT UPTAKES AND CORN YIELD

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Abstract

Phosphorus use efficiency in soils of Pakistan is reported less than 15%. However, phosphorus (P) bio-fertilizers have potential to improve P use efficiency (PUE) in these soils on sustainable basis. A primary study was conducted under control conditions of greenhouse to compare P availability from reduced rates of phosphorus bio-fertilizer (PSB-DAP) with P available from 100% rate of DAP recommended for maize. Based on the findings, a field study was conducted to evaluate the effects of 100% rate of uncoated DAP and 50-75% rate of PSB-DAP on corn yield and PUE. The treatment for both studies included; control (with fertilizers), 100% of recommended P₂O₅ from Commercial DAP (C-DAP₁₀₀), 75% of recommended P₂O₅ from PSB-DAP (PSB-DAP₅₀) and 50% of recommended P₂O₅ from PSB-DAP (PSB-DAP₅₀). Results explored that available P from PSB-DAP₅₀ and C-DAP₁₀₀ was statistically at par while PSB-DAP₇₅ at 30, 45 and 60 days of incubation. The findings of field study also revealed that PBS-DAP improved cob weight by 5-10%, chlorophyll contents by 14-23%, 1000-grains weight by 8-10%, grain yield by 10%, and P uptake by 35%, PUE efficiency by 42% compared to C-DAP₁₀₀. This work can be used as a case study to promote P bio-fertilizers among farming community and researchers in order to for make effective use of alternate P sources for crop production

Keywords: PSB, biofertilizer, impregnated DAP, PUE, maize.

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1. INTRODUCTION

Maize is one of the most important crop and globally known as queen of cereals (Gong et al., 2024). There is a continuous increase in the gaps between potential yield and actual productivity of maize. Better phosphorus (P) management practices can be used to exploited through the adoption of hybrids (Ciampitti and Vyn, 2014). Phosphorus is the second most important macronutrient after nitrogen (N) (Pandey, 2018). Its sufficient supply is essential for seedling growth, early rooting and maturity, formation and quality of seed and photosynthetic water use efficiency

(Memon et al., 2011). It is also required for ATP biosynthesis during respiration and photosynthesis and energy transfer (Reddy et al., 2002). Thus, soil deficient in P is a major challenge because 80% of applied P from synthetic fertilizers immediately either precipitates or converts to plant unavailable P (Fernandez et al., 2007). Phosphorus use efficiency (PUE) is poor in soils featured with calcareousness, alkaline pH and deficient organic matter. Most of cultivated soils of Pakistan (90%) are P deficient because 75-85% of applied P is lost (Memon et al., 2011). Therefore, phosphorus management always remained



This work is licensed under a <u>Creative Commons</u> <u>Attribution-NonCommercial 4.0 International License</u> among first preferences while considering the macronutrients supply to the crops (Salazar et al., 2011, Pandey, 2018). However, recent development in soil microbiology is expected to resolve the low PUE in such soils.

Soil microbes especially phosphorus bacteria (PSB) play a solubilizing significant role in phosphorus dynamics of alkaline or calcareous soils. These PSB prolong P availability to plant roots in soils by two mechanisms; 1) solubilize the fixed P through organic acids and 2) avoid precipitation of Ca⁺² and Mg⁺² in alkaline and calcareous soils (Afzal and Bano, 2008). Moreover, the PSB lowers pH to stop the adsorption of P on calcium carbonate surface. In this way, PSB may play critical roles in sustainable agriculture models (Estrada-Bonilla et al., 2017).

The role of P fertilizers for cereal crops throughout the world cannot be ignored. Maize is one of the key cereal crop after wheat and rice. It provides proteins, (10%), carbohydrates (72%), oils (4.8%), fiber (5.8%) and mineral. In calcareous soils, hybrid cultivars of maize fail to give their potential yield due to poor PUE and thus per acre maize yield in Pakistan is too much lower than important maize producers. Arif et al. (2018) reported a significant improvement in P physiological and recovery efficiency when they incubated seeds with PSB. Keeping in view all above discussion and based on the findings of Arif et al. (2018), the research was conducted to evaluate the effects of PSB coated DAP with 25-50% of recommended P for improving PUE and yield of maize.

2. Materials and Methods

2.1. Experimental site

Lab study was conducted in the laboratory of Department of Soil and Environmental Sciences while field study was carried out at the research area of MNS University of Agriculture Multan, Pakistan (30°08'51"N 71°26'36"E, 122.19 m) during 2017-2018.

2.2. Experimental soil characteristics The soil samples (0-15 cm depth) from the experimental site were collected and analyzed (Table 1). The soil used was of loam texture. The soil of experimental site was of loam texture with deficient nutritional and organic matter status and alkaline in nature (Table 1). The soil from the same experimental field was used during pot study.

Table 1: Experimental field's physico-chemical soil characteristics

Parameter	Unit	Value			
Textural class	-	Loam			
Organic matter	%	0.49			
ECe	dS m^{-1}	2.11			
pН	-	8.00			
CEC	Cm _c kg ⁻¹	5.29			
Total N	%	0.039			
Available P	mg kg ⁻¹ soil	6.5			
Extractable K	mg kg ⁻¹ soil	110			

2.3. Lab study

The treatment plan of lab study included control (soil without DAP, T₁), 100% of recommended P2O5 from commercial DAP (T_2) and 75% of recommended P₂O₅ from PSB inoculants (PSBI) and three incubation periods (30, 45 and 60 days of incubation). The lab study was laid down using two factorial CRD with four replications. Fine ground soil (100 g cup⁻¹) taken from experimental field was mixed with FFC commercial DAP $(1.00 \text{ g cup}^{-1})$ and polymer coated DAP (0.75 g cup⁻¹). The DAP mixed soil was filled in disposable cups and placed in an incubator (Syano Incubator) for incubation up to 60 days. Water required for soil saturated paste was measured and 75% of the calculated water (equivalent to field capacity) was added to each cup and weighed. Later on, the moisture in the soil of each cup was maintained on weight basis.

2.4. PSB isolation and inoculation on DAP grains

University of Agriculture, Faisalabad, Pakistan provided a pre-isolated strain (P-10) of PGPR containing ACC-deaminase and phosphatase activity. The strain was multiplied using general purpose medium (GPM) in the conical flasks (250 mL) and used for coating over diammonium phosphate (DAP) using methods discussed by Arif et al. (2018) for coating of PSB on single superphosphate (SSP).

2.5. Field study

Like lab study, a field study was also conducted to evaluate the effects of PSB coated DAP on nutrient use efficiency, growth and yield of maize. Field study was laid out according to randomized complete block design (RCBD) with four replicate. The detail of treatment plan was: $T_1 =$ Control (without P₂O₅ application; DAP₀), $T_2 = 100\%$ of recommended P_2O_5 from DAP (DAP₁₀₀), $T_3 = 75\%$ of recommended P₂O₅ from PSB coated DAP (PSB-DAP₇₅) and $T_4 = 50\%$ of recommended P_2O_5 from PSB coated DAP (PSB-DAP₅₀). In all treatments, N and K₂O were applied @ 150 and 90 kg ha⁻¹ whereas the recommended rate for P_2O_5 was 120 kg ha⁻¹.

2.5.1. Land preparation, treatment application, sowing and data collection

The seedbed was prepared after ploughing and planking. For bed preparation, seedbed planter was used. On each side of bed (2.5 feet wide), seeds of maize was sown manually while maintaining plant-to-plant distance 2 feet. The source of N and K used were Urea and SOP, respectively. All the amount of P and K fertilizers was added before sowing and N was added in three splits i.e. before sowing, at first irrigation and at fourth irrigation. The field was irrigated at Field capacity. For weed control, pre-emergence "Pendimethalin" was used while post-emergent weeds were eradicated manually. For control of sucking and chewing insects, Syngenta company products were used after pest scouting.

The data related to growth attributes was taken before harvesting of crop while yield attributes was taken after harvest of crop. different Similarly, data related physiological such parameters as photosynthetic rate. transpiration, chlorophyll contents was collected using Portable Chlorophyll meter SPAD-502 and IRGA (Infra-Red Gas Analyzer) as discussed in Ahmed et al. (2020).

2.6. Fertilizer uptakes and use efficiency

Following procedure given by Wolf (1982), Jackson (1962) and Chapman and Pratt (1961), N, P and K concentration in shoot and corn grains were determined. The collected plant samples and grains were dried in Air flow oven at 65°C till constant weight, ground and stored in a plastic bag till wet digestion. The wet digestion was done using mixture of hydrogen peroxide and sulphuric acid (Ryan et al., 2001). From digested samples, N was measured using Kjeldhal and distillation method whereas P was measured by following vanadium phosphospectrophotometric molybdate method (Olsen et al., 1954). K content from digested samples was measured using flame photometer (Chapman and Pratt, 1961). N, P, K uptakes were calculated as product of N, P and K concentration and dry weight plant⁻¹, respectively while different forms of nutrient use efficiency were also calculated using following formulae used by Ahmed et al. (2020):

$$AUE (kg kg^{-1}) = \frac{GY_T - GY_C}{NA}$$
$$AR(\%) = \frac{NU_T - NU_C}{NA} \times 100$$
$$PUE (kg kg^{-1}) = \frac{GY_T - GY_C}{NU_T - NU_C}$$
$$IR (\%) = \frac{NU_T - NU_C}{NU_T} \times 100$$
$$UTE(\%) = \frac{GY_C}{GY_T} \times 100$$
$$FSF(\%) = \frac{GY_T - GY_C}{GY_T} \times 100$$

Whereas AUE = agronomic use efficiency, GYT = Grain yield in treated plot, GYc =Grain yield in control plot, NUT = nutrient uptake in treated plot, NUc = nutrient uptake in control plot, NA = nutrient applied, AR = Apparent recovery, PUE =Physiological use efficiency, IR = Internal remobilization, UTE Utilization =efficiency and FSF = Fertilizer stress factor. Physiological use efficiency, IR = Internal remobilization, UTE = Utilization efficiency and FSF = Fertilizer stress factor. **Statistical analysis**

All the data was analysed using completely randomized design (CRD) for two-way analysis of variance (ANOVA) in lab study while randomized complete block design (RCBD) for one-way ANOVA in field study. The difference in mean values were compared using LSD test at 5% probability level (Steel et al., 1997). The data was analysed statistically using Statistix 8.1©.

3. Result and Discussion:

3.1. Variations in available P under controlled conditions

The data related to available P from PSB-DAP (75%) and commercial DAP (100%) at 30, 45 and 60 days of incubation is shown in Table 2. Results of Table 2 showed a gradual increase in available P with the passage of time. At 60 days of incubation, Olsen P was found at higher from PSB-DAP and commercial DAP. Results show that the Olsen P from 1.00 g commercial DAP recommend P from PBS-DAP and 0.75 g PBS-DAP was statistically at par (Table 2). This might be happened due to the physiological role of PBS strain coated over DAP. As the soil used was alkaline and deficient in organic matter, the P released from the commercial DAP was immediately fixed in the soil where PSB strain was not used. Arif *et al.* (2018) also used the same strain of PSB but coated over single super phosphate (SSP) and reported significant improvements in P uptake and use efficiency in cotton.

3.2. Plant biomass and chlorophyll contents

The findings of field study clearly depict that plants fertilized with PSB-DAP had higher plant dry mass than those plants fertilized with commercial DAP (Fig. 1). Data related to plant dry biomass also reveal that the effect of 100% P from commercial DAP and 50% P from PSB-DAP. However, chlorophyll contents

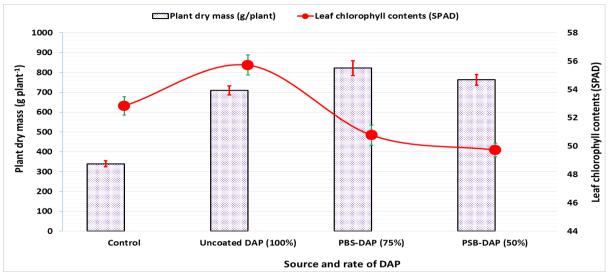


Fig. 1. Comparative effect of P from uncoated DAP and PSB-DAP on plant dry mass and chlorophyll contents of maize. Note: PSB-DAP = PSB coated DAP

Table 3. Variations in growth	variables of ma	ize due to	polymer	coated	DAP	and
uncoated DAP under field condi	tions					

Treatment	Cob	1000-	Grain yield		Water	Use Efficiency	
	weight (g cob ⁻¹)	grains weight (g)	g plant ⁻¹	tons acre ⁻¹	Plant WUE	Photosynthetic WUE	
Control	136 c	235 c	108 c	1.886 c	18.6 c	0.37 c	
DAP ₁₀₀	284 b	423 b	248 b	4.312 b	41.3 b	0.45 b	
PSB-DAP75	329 a	490 ab	273 a	4.748 a	45.6 a	0.71 a	
PSB-DAP ₅₀	305 ab	503 a	250 b	4.356 b	46.3 a	0.77 a	
LSD value	17.740	92.462	21.073	0.30217	12.614	0.2952	

Values sharing same letter(s) in each column do not differ by each other at p = 0.05 according to LSD test. Plant WUE (g/mmol m⁻² s⁻¹), Yield per acre acre acre values according to 17242 plants per acre.

declined in plants where P source was PSB-DAP. This decline in chlorophyll content might be due to lower supply of N from PSB-DAP₇₅ and PSB-DAP₅₀. But contrary to chlorophyll contents, improvement in plant dry mass either due to PSB-DAP₇₅ or PSB-DAP₅₀ might be occurred due to more supply of P to plant roots as it was observed in lab study that PSB showed positive effect on Olsen P.

3.3. Grain yield attributes and water use efficiency

We observed that the plants having supply of P from PSB-DAP₇₅ and PSB-DAP₅₀ improved cob weight, 1000-grains weight and yield per plant compared to yield of plants treated with 100% P from uncoated DAP (Table 3). Significant improvement in cob weight and 1000-corn grains was also recorded due to the dynamic role of PSB. The PSB not only reduced the DAP rate of application but also resulted in 10% more grain yield. This might be happened due to physiological role of P in grain and root development. Like yield per plant, similar improvements were also found in water use efficiency (WUE). We also noticed a gradual increase in both types of WUE; Plant water use efficiency and photosynthetic water use efficiency due to P supply from PSB-DAP₇₅ and PSB-DAP₅₀ as compared 100% P supply from uncoated DAP (T₂). Arif et al. (2018), Yaseen et al. (2018) and Billah et al. (2015) also reported improvement in yield attributes cotton and microbial wheat due to activity. Improvements in grain weight and water use efficiency occurred might be due to sufficient P supply which is essential for photosynthetic maturity. WUE and formation and quality of seed (Memon et al., 2011).

3.4. Plant NPK contents

In calcareous soils, nutrient uptake is of greater concern. Nutrient losses in these soils are higher i.e. 60-90, 90, 60 and 60% losses of N, P, K and other nutrients, respectively. We used the sum of shoot and grain N contents (mg NPK plant-1) as an approximate measure of total N uptake by

Treatment	NPK concentration (mg g ⁻¹)		NPK contents (mg plant ⁻¹)*			
	Ν	Р	K	Ν	P	K
Control (T ₁)	7.1 b	3.65 c	6.4 c	32 c	0.39 c	33 c
$DAP_{100}(T_2)$	10.2 a	4.21 b	8.0 b	90 a	1.04 b	45 c
$PSB-DAP_{75}(T_3)$	7.7 b	4.52 a	10.4 a	60 b	1.23 a	91 a
PSB-DAP $_{50}(T_4)$	8.7 b	4.36 b	9.8 a	50 b	1.09 b	68 b
LSD values	2.2044	0.1533	1.1731	25.103	0.1725	16.367

Table 4. Comparative effects of PBS-DAP and uncoated DAP on plant N, P and K concentration and uptake in maize under alkaline/ calcareous conditions

Values sharing same letter(s) in each column do not differ by each other at p = 0.05 according to LSD test. *NPK uptake

Table 5. Comparison amo	ng different form	s of PUE in maize grov	vn on calcareous soil
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Treatment	PAUE (g g ⁻¹)	PRE (%)	PPUE (g g ⁻¹)	IPR (%)	PUTE (%)	PSF (%)
Control (T ₁)	-	-	-	-	-	-
$DAP_{100}(T_2)$	37 c	17 c	215 a	62 c	44 b	56 a
$PSB-DAP_{75}(T_3)$	59 b	30 b	197 c	68 a	40 b	40 b
$PSB-DAP_{50}(T_4)$	76 a	37 a	204 b	64 b	43 a	43 b
LSD values	12.614	4.132	5.234	1.042	2.2044	10.422

Values sharing same letter(s) in each column do not differ by each other at p = 0.05 according to LSD test. P agronomic use efficiency (PAUE), P recovery efficiency (PRE), P physiological use efficiency (PPUE), Internal P remobilization (IPR), P utilization efficiency (PUTE) and P stress factor (PSF)

plant. We noted the greatest N uptake in plants subjected to treatment T4 while the least N uptake in plants of control treatment without application of fertilizers (Table 4). Table 3 also suggests that the P uptake by plant varies with P supply source to plants. Our calculations explore that the P uptake is dependent on P source. The P released from peaks in plants subjected to treatment T3 (Table 4). In contrast, P coming out from DAP resulted in a decrease in P uptake by plant. In nutshell, we suggest the usage of PSB based fertilizers in calcareous soil for effective P uptake by plants. The P uptake by plants significantly affected plant biomass. The plants subjected to treatment T₂ showed the greatest N uptake while the plants of control treatment had the least N and K uptake (Table 4). The plants with greater P uptakes also expressed greater K uptake and vice versa (Table 4). Yaseen et al. (2018) also reported similar findings.

3.5. Phosphorus use efficiency

Maize plants subjected to different DAP fertilizers responded differently for PUE. Plants subjected to treatment T₄ (PSB-DAP₅₀) had the highest PAUE (204 g grain/g P applied), and PRE (37%) and PUTE (43%), followed by T₃ (PSB-DAP₇₅) (Table 5). Alike, Table 5 also reveal that the plants subjected to PSB-DAP₇₅ showed the highest IPR (68%). We suggest that this higher PRE is due to higher P uptake by the plants supplied with PSB-DAP (Table 4). The decline in PPUE in treatment T₃ (PSB-DAP₇₅) and treatment T₄ (PSB-DAP₅₀) i.e. 197 g grain/g and 204 g grain/g P applied, respectively, were observed compared to treatment T₂ i.e. 215 g grain/g P applied (Table 5). This decline in PPUE suggests effective conversion of absorbed P into grains or economical portion of maize. The plants subjected to different forms of DAP behave contrarily for P utilization efficiency (PUTE) and P stress factor (PSF). The treatment PSB-DAP₇₅ had the lowest PUTE (40%) compared to uncoated DAP (44%) whereas PSF in plants treated with PSB-DAP₇₅ was 40% compared to 56% PSF in plants subjected to uncoated

DAP, indicating that PSB kept P in available form and enhanced P uptake (Table 4). These improvements in P use efficiency occurred might be due to physiological role of P and effective P uptake in plants. Arif *et al.* (2018), Yaseen *et al.* (2018) and Billah *et al.* (2015) also reported similar findings in PUE in cotton and wheat respectively due to PSB. Improvements in yield attributes occurred might be due to sufficient P supply which is essential for maturity, photosynthetic WUE and formation and quality of seed (Memon *et al.*, 2011).

4. Conclusion

PSB coated DAP (Phosphatic bio fertilizer) caused a significant increase in plant biomass, chlorophyll contents, water and P use efficiency and ultimately corn grain yield even after application of 25 to 50% P from recommended P for maize. This study suggests that P biofertilizer performed better under reduced application of 50% of the recommended P. It also suggests that PSB coated DAP has a great potential to cut down pressure on chemical fertilizers and P exhausting resources. Based on the findings of these studies, researchers and breeders can develop such P responsive cultivars, having potential to utilize major portion of unavailable P.

5. Conflict of Interest

There is no conflict of interest among the authors related to this work.

6. Acknowledgement:

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7. Author's Contribution:

All the authors contributed in their domains related to lab and field study.

8. REFERENCES

Afzal, A., and A. Bano, 2008. Rhizobium and phosphate solubilizing bacteria improve the yield and phosphorus uptake in wheat (*Triticum aestivum*). Int. J. Agric. Biol.10:85-88.

- Ahmed, W., M. Imran, M. Yaseen, T. ul Haq, M.U. Jamshaid, S. Rukh, and M.A. Khan. 2020. Role of salicylic acid in regulating ethylene and physiological characteristics for alleviating salinity stress on germination, growth and yield of sweet pepper. PeerJ 8:e8475.
- Arif, M., W. Ahmed, U. Jamshaid, M. Imran and S. Ahmad. 2018. Effect of rock phosphate based compost and biofertilizer on uptake of nutrients, nutrient use efficiency and yield of cotton. Soil Environ. 37:129-135.
- Billah, M. and A. Bano. 2015. Role of plant growth promoting rhizobacteria in modulating the efficiency of poultry litter composting with rock phosphate and its effect on growth and yield of wheat. Waste Manag. Res. 33:63-72.
- Chapman, H. D. and F.P. Pratt. 1961. Ammonium vandate-molybdate method for determination of phosphorus. Methods Anal. Soils, Plants Water 1:184-203.
- Ciampitti, I.A. and T.J. Vyn. 2014. Understanding global and historical nutrient use efficiencies for closing maize yield gaps. Agron. J. 106(6): 2107-2117.
- Estrada-Bonilla, G.A., C.M. Lopes, A. Durrer, P.R. Alves, N. Passaglia and E.J. Cardoso. 2017. Effect of phosphate-solubilizing bacteria on phosphorus dynamics and the bacterial community during composting of sugarcane industry waste. Syst. Appl. Microbiol. 40:308-313.
- Gong, H., Y. Xiang, J. Wu, L. Luo, X. Chen, X. Jiao and C. Chen. 2024. Integrating phosphorus management and cropping technology for sustainable maize production. J. Integrative Agric. 23:1369-1380. Jackson, M.L. 1962. Chemical composition of soil. pp. 71-144. *In:* Chemistry of Soil. Bean

F.E. (ed.). Van -Nostrand Co., New York. USA.

- Memon, M. S., J.A. Shah, P. Khan, M. Aslam and N. Depar. 2011. Effect of phosphorus fertigation in wheat on different soils varying in CaCO3 levels. Pak. J. Bot. 43:2911-14.
- Pandey, N. 2018. Role of plant nutrients in plant growth and physiology. Plant Nutr. Abio. Str. Tol. 51-93.
- Reddy, M. S., S. Kumar, K. Babita and M.S. Reddy. 2002.
 Biosolubilization of poorly soluble rock phosphates by Aspergillus tubingensis and Aspergillus niger. Biores. Tech. 84:187-189.
- Ryan, J., G. Estefan and A. Rashid. 2001. Soil and Plant Analysis Laboratory Manual. 2nd Edition. ICARDA. pp. 141-155.
- Salazar, S., L.E. Sánchez, J. Alvarez, A. Valverde, P. Galindo, J.M. Igualand and I. Santa-Regina. 2011. Correlation among soil enzyme activities under different forest system management practices. Ecol. Engin. 37:1123-1131.
- Steel, R.G.D., J.H. Torrie and D.A. Dicky. 1997. Principles and procedures of statistics. A biological approach. p. 352-358. 3rd Ed. McGraw Hill. Inc. Book Co. New York, USA.
- Wolf, B. 1982. The comprehensive system of leaf analysis and its use for diagnosing crop nutrient status. Commun. Soil Sci. Plant Anal. 13:1035-1059.
- Yaseen, M., A. Abbas, M.Z. Aziz, A. Wakeel, H. Yasmeen, W. Ahmad and Naveed. 2018. Microbial assisted foliar feeding of micronutrients enhance growth, yield and biofotification of wheat. Int. J. Agric. Biol. 20:353-360.